mNeongreen Nanoselector Agarose

Summary

Catalog No	013-101-002
Ligand	Anti-mNeongreen single domain antibody fragment (VHH, Nanobody)
Bead size	~ 40 µm
Reactivity	Recognizes mNeongreen selectively
Binding capacity	High binding capacity, 10 µL slurry bind about 20 µg of recombinant mNeongreen
Storage	Shipped at ambient temperature. Upon receipt store at 4°C. Stable for 1 year. Do not freeze
Storage buffer	50 % slurry in PBS containing 20 % Ethanol

Description

mNeongreen Nanoselector Agarose has been specifically designed to bind mNeogreen-fusion proteins. mNeongreen Nanoselector Agarose is based on small high-affinity recombinant single domain antibody covalently coupled to the surface of Agarose beads. mNeongreen Nanoselector Agarose is an ideal tool to isolate or purify mNeongreen-fusion proteins fast and efficiently.

Background

mNeongreen is the brightest monomeric green or yellow fluorescent protein yet described to our knowledge, performs exceptionally well as a fusion tag for traditional imaging as well as stochastic single molecule super-resolution imaging and is an excellent fluorescence resonance energy transfer(FRET) acceptor for the newest cyan fluorescent proteins. With the mNeongreen Nanoselector, specific for this non-GFP variant, yields of fusion proteins purification are high and free from heavy and light-chain contaminants, all in under 30 minutes. Save time, save effort and get cleaner results. The mNeongreen Nanoselector is the use of super-high affinity Alpaca antibody fragments called nanobodies. mNeongreen Nanoselector may be used for immuno-precipitation, immuno-purification and immuno-pull down experiments with up to 10-fold better purity and yield than that of classic mouse monoclonal antibodies. Compatible with a variety of source materials, Nanoselector may be used with mammalian cells, tissues & organs, bacteria, yeast and even plants. These reagents allow your mNeogreen-fusions to be perfect candidates for immune-precipitations, Co-IP, mass spectroscopy, and enzyme activity measurements.

Application notes

Immunoprecipitation (IP)/Co-IP
Mass spectrometry (MS)
Enzyme activity measurements

Benefits

- Effective pulldown of mNeongreen-fusion proteins for consistent results
- No heavy & light antibody chains, short incubation (5-30 min)
- Extraordinary binding, even under harsh conditions
- Very high affinity to bind even low abundant proteins

Immunoprecipitation protocol

Mammalian cell lysis

Note: Harvesting of cells and cell lysis should be performed with ice-cold buffers. We strongly recommend to add protease inhibitors to the Lysis buffer to prevent degradation of your target protein and its binding partners.

For one immunoprecipitation reaction, we recommend using ~106- 107 cells.

1. Choice of lysis buffer:

* For cytoplasmic proteins, resuspend the cell pellet in 200 µL ice-cold Lysis buffer by pipetting up and down. Supplement Lysis buffer with protease inhibitor cocktail and 1 mM PMSF (not included).
* For nuclear/chromatin proteins, resuspend cell pellet in 200 µL ice-cold RIPA buffer supplemented with DNaseI (f.c. 75-150 Kunitz U/mL), MgCl2 (f.c. 2.5 mM), protease inhibitor cocktail and PMSF(f.c. 1 mM)(not included)

2. Place the tube on ice for 30 min and extensively pipette the suspension every 10 min.
3. Centrifuge cell lysate at 17,000x g for 10 min at +4°C. Transfer cleared lysate (supernatant) to a pre cooled tube and add 300 µL Dilution buffer supplemented with 1 mM PMSF and protease inhibitor cocktail (not included). If required, save 50 µL of diluted lysate for further analysis (input fraction).

Bead equilibration

1. Resuspend the beads by gently pipetting up and down or by inverting the tube. Do not vortex the beads!
2. Transfer 25 µL of bead slurry into a 1.5 mL reaction tube.
3. Add 500 µL ice-cold Dilution buffer.
4. Sediment the beads by centrifugation at 2,500x g for 5 min at +4°C.
5. Discard the supernatant.

Protein binding

1. Transfer 25 µL of equilibrated beads to the tube of protein binding.
2. Rotate end-over-end for 1 hour at +4°C.

Washing

1. Sediment the beads by centrifugation at 2,500x g for 5 min at +4°C.
2. If required, save 50 µL of supernatant for further analysis(-flow-through/non-bound fraction).
3. Discard remaining supernatant.
4. Resuspend beads in 500 µL Wash buffer.
5. Sediment the beads by centrifugation at 2,500x g for 5 min at +4°C. Discard the remaining supernatant.
6. Repeat this step at least twice.
7. During the last washing step, transfer the beads to a new tube.

Optional: To increase stringency of the Wash buffer, test various salt concentrations e.g. 150 mM - 500 mM,and/or add a non-ionic detergent e.g. Triton™ X-100.

Elution with 2x SDS-sample buffer

1. Remove the remaining supernatant.
2. Resuspend beads in 80 µL 2x SDS-sample buffer.
3. Boil beads for 5 min at +95°C to dissociate immunocomplexes from beads.
4. Sediment the beads by centrifugation at 2,500x g for 2 min at +4°C.
5. Analyze the supernatant in SDS-PAGE.

Elution with Glycine-elution buffer

1.Remove the remaining supernatant.
2. Add 50–100 µL Glycine-elution buffer and constantly pipette up and down for 30 - 60 sec at +4°C.and down for 30 - 60 sec at +4°C.
3. Sediment the beads by centrifugation at 2,500x g for 5 min at +4°C.
4. Transfer the supernatant to a new tube.
5. Immediately neutralize the eluate fraction with Neutralization buffer.
6. Repeat this step at least once to increase elution efficiency .

Suggested buffer compositions

Buffer	Composition
Lysis buffer	10 mM Tris/Cl pH 7.5, 150 mM NaCl, 0.5 mM EDTA, 0.5 % NP40
RIPA buffer	10 mM Tris/Cl pH 7.5, 150 mM NaCl, 0.5 mM EDTA, 0.1 % SDS, 1 % Triton™ X-100, 1 % deoxycholate
Dilution/Wash buffer	10 mM Tris/Cl pH 7.5, 150 mM NaCl, 0.5 mM EDTA
2x SDS-sample buffer	120 mM Tris/Cl pH 6.8, 20 % glycerol, 4 % SDS, 0.04 % bromophenol blue, 10 % β-mercaptoethanol
Glycine-elution buffer	200 mM glycine pH 2.0
Neutralization buffer	1 M Tris pH 10.4

Related products

Code Number	Product Description	Size	prices(¥)
019-101-002	GFP Nanoselector Agarose	0.25mL	1500
019-101-003	GFP Nanoselector Magnetic beads	0.25mL	1500
020-101-002	RFP Nanoselector Agarose	0.25mL	1500
020-101-003	RFP Nanoselector Magnetic beads	0.25mL	1500
013-101-002	mNeongreen Nanoselector Agarose	0.25mL	1500
014-101-002	TurboGFP Nanoselector Agarose	0.25mL	1500
015-101-002	MBP Nanoselector Agarose	0.25mL	1500
010-101-002	GST Nanoselector Agarose	0.25mL	1500
011-101-002	SNAP tag Nanoselector Agarose	0.25mL	1500
012-101-002	Halo Nanoselector Agarose	0.25mL	1500
003-101-002	HA tag Nanoselector Agarose	0.25mL	1500
004-101-002	c-His tag Nanoselector Agarose	0.25mL	1500
049-101-002	mWasabi Nanoselector Agarose	0.25mL	1500
017-101-002	TagFP Nanoselector Agarose	0.25mL	1500
025-101-002	Rabbit IgG Nanoselector Agarose	0.25mL	1500
001-101-002	Mouse IgG Nanoselector Agarose	0.25mL	1500
067-101-003	Streptavidin Magnetic beads	0.25mL	1500
100-100-100	Binding Control Nanoselector Agarose	1mL	800
100-100-200	Binding Control Magnetic beads	1mL	800